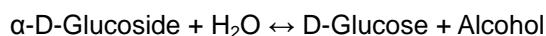


α -GLUCOSIDASE (α -Glu)

[EC 3.2.1.20]

from *Bacillus stearothermophilus*



SPECIFICATION

State	: Lyophilized	
Specific activity	: more than 40 U/mg protein	
Contaminants	: (as α -Glu activity = 100 %)	
	Phosphoglucomutase	< 0.01 %
	NADH oxidase	< 0.01 %
	Alcohol dehydrogenase	< 0.01 %

PROPERTIES

Molecular weight	: ca. 50,000	
Optimum pH	: 6.0 - 7.0	(Fig. 1)
pH stability	: 5.0 - 11.0	(Fig. 2)
Isoelectric point	:	
Thermal stability	: No detectable decrease in activity up to 60 °C.	(Fig. 3, 4)
Michaelis constants	: (50 mM Potassium phosphate buffer, pH 6.3, at 30 °C)	
	p-Nitrophenyl- α -glucopyranoside (PNPG)	0.73 mM
	Maltose	1.3 mM
	Phenyl- α -glucopyranoside	2.4 mM
Substrate specificity	: PNPG	100 %
	Maltose	177 %
	Phenyl- α -glucopyranoside	59 %

STORAGE

Stable at -20 °C for at least one year

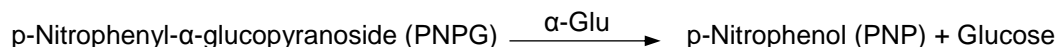
APPLICATION

The enzyme is useful for diagnostic reagent, for example, α -amylase determination.

ASSAY

Principle

The change in absorbance is measured at 400 nm according to the following reaction.



Unit Definition

One unit of activity is defined as the amount of α -Glu that forms 1 μmol of PNP per minute at 30 °C.

Solutions

- I Buffer solution ; 100 mM Potassium phosphate buffer, pH 6.3
- II PNPG solution ; 20 mM (0.603 g PNPG/100 mL distilled water) (Stable for two weeks if stored at 0 - 5 °C)
- III Na_2CO_3 solution ; 0.2 M (2.12 g Na_2CO_3 /100 mL distilled water)

Preparation of Enzyme Solution

Dissolve the lyophilized enzyme with distilled water and dilute to 0.006 to 0.022 U/mL with 10 mM Potassium phosphate buffer containing 1 mg/mL BSA, pH 7.5.

Procedure

1. Prepare the following reaction mixture and pipette 1.5 mL of reaction mixture into a test tube.
Solution I 10.0mL
Solution II 5.0mL
2. Incubate at 30 °C for 5 minutes.
3. Add 0.5 mL of the enzyme solution and mix.
4. Incubate at 30 °C for exactly 15 minutes.
5. After incubation, add 2.0 mL of Solution III and mix.
6. Read absorbance at 400 nm (Abs•test).
At the same time, prepare the blank with 1.5 mL of the reaction mixture, and add 2.0 mL of Solution III after incubation at 30 °C for 15 minutes, followed by addition of the enzyme solution (Abs•blank).

Calculation

$$\text{Volume activity (U/mL)} = ((\text{Abs}\cdot\text{test}) - (\text{Abs}\cdot\text{blank})) \times \frac{4.0}{18.1 \times 15 \times 0.5} \times \text{d.f.}$$

$$\text{Specific activity (U/mg protein)} = \frac{\text{Volume activity (U/mL)}}{\text{Protein concentration (mg/mL)}^*}$$

d.f. ; dilution factor

18.1 ; millimolar extinction coefficient of PNP ($\text{cm}^2/\mu\text{mol}$)

*Protein concentration ; determined by Bradford's method

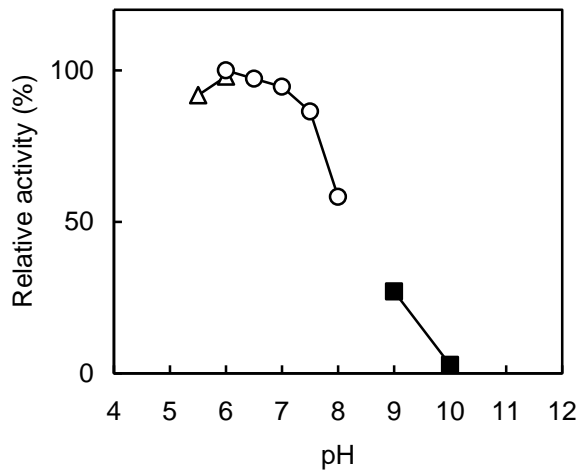


Fig. 1 pH profile

(Δ acetate, \circ phosphate,
 \blacksquare Gly-NaOH)

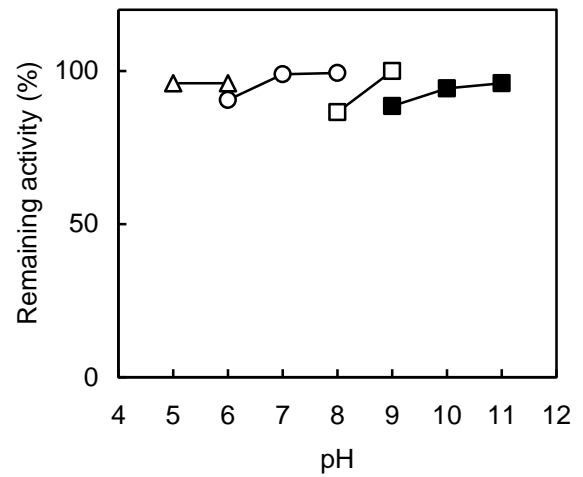


Fig. 2 pH stability

(treated for 24 hr at 4 °C in the following buffer solution (0.1 M);
 Δ acetate, \circ phosphate,
 \square TEA-NaOH, \blacksquare Gly-NaOH)

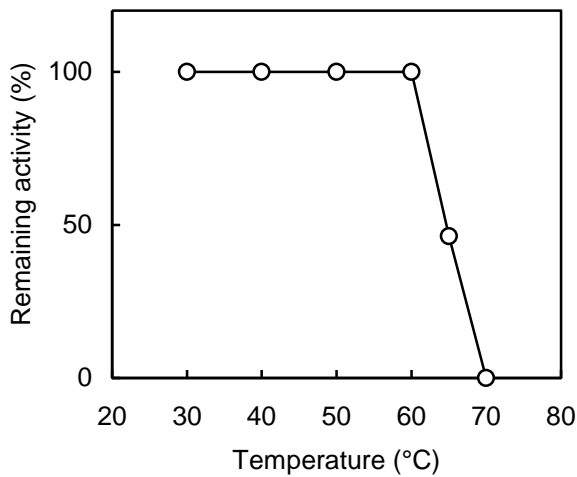


Fig. 3 Thermal stability

(treated for 15 min in 0.1M potassium phosphate buffer, pH 8.0)

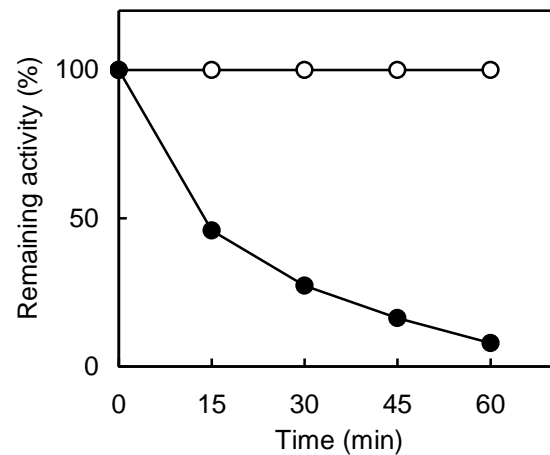


Fig. 4 Thermal stability

(treated for in 0.1M potassium phosphate buffer, pH 8.0
 \circ 60 °C, \blacksquare 65 °C)